

POSTER PRESENTATION

Open Access

Profiling of glycosylation gene expression in CHO fed-batch cultures in response to glycosylation-enhancing medium components

Ryan Boniface^{1*}, Jeoffrey Schageman², Brian Sanderson², Michael Gillmeister¹, Angel Varela-Rohena¹, John Yan³, Yolanda Tennico³, Shawn Barrett¹, Robert Setterquist², Stephen Gorfien¹

From 23rd European Society for Animal Cell Technology (ESACT) Meeting: Better Cells for Better Health Lille, France. 23-26 June 2013

Introduction

Characterization of the glycosylation profile of a recombinant protein product is an important part of defining product quality in the bioproduction industry. Development of a protein with desired characteristics would require the capacity to modify and target specific glycosylation patterns as well as an understanding of the implications of changes to these glycosylation profiles. Previous cell culture studies have demonstrated the ability to modulate glycan profiles without negative impact to culture growth and product titer through the addition of glycosylation-enhancing medium components. With new methods, including increased measurement sensitivity and new capabilities in RNA-Seq technology, it is possible to develop a glycosylation gene expression profile for CHO cells. Specific glycosylation genes can then be tracked to ensure that the addition of these compounds will not negatively impact gene expression. Analyses comparing growth and titer, glycan distribution, and transcriptome differences can present us with potential insight into what changes are taking place on a genetic level in the cell in response to changes in medium and culture conditions.

Materials and methods

(All Materials were from Life Technologies unless otherwise indicated)

Cell culture

CHO-S[®] and DG44 derived recombinant cells expressing the same IgG molecule were grown in CD FortiCHO[™]

medium supplemented with 4mM L-glutamine and 1:100 Anti-Clumping Agent.

Fed-batch bioreactor

DASGIP bioreactor with 500mL initial working volume seeded at 0.3×10^5 viable cells/ml in CD FortiCHO[™] medium. 10% CD EfficientFeed[™] C (EFC) feeding on days 3, 5 and 7 for CHO-S[®] cultures, and feeding on days 4, 6 and 8 for DG44 cultures. Glucose concentration was maintained above 3g/L. Component A and/or component B were added on the first day of feeding (day 3 for CHO-S[®] and day 4 for DG44 cultures). Culture conditions were maintained as follows; pH 7.0 \pm 0.05, 50% DO, 37°C, 110 rpm. Cell densities and viabilities were measured using a Vi-CELL[®] counter (Beckman Coulter). Metabolites (glucose, ammonia, lactate) and IgG were measured using a Cedex[®] Bio HT Instrument (Roche).

Glycan analysis

Protein supernatant samples were collected and purified using POROS[®] MabCapture[®] A resin. Samples glycan profiles were analyzed on an Applied Biosystems[®] 3500 Series Genetic Analyzer.

Transcriptome analysis

RNA was extracted at several time points during the culture. A total of 174 potential glycosylation specific gene targets were identified and primers designed to these using reference sequences from Chinese hamster ovary, mouse, rat and human. A total of 34 samples were multiplexed on a Proton[™] PI chip on the Ion Torrent[™] PGM[™].

* Correspondence: Ryan.Boniface@lifetech.com

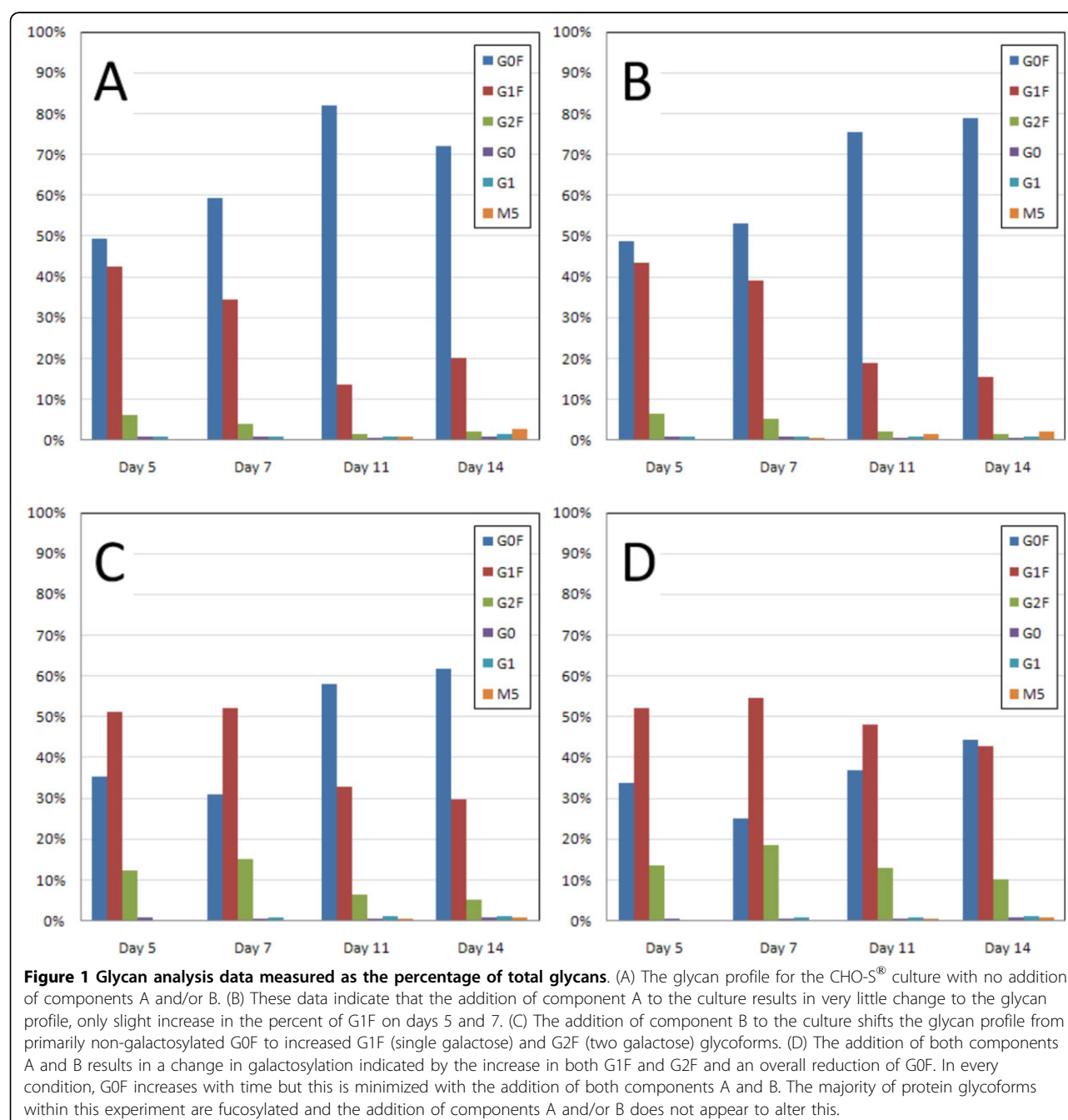
¹Life Technologies Corporation, 3175 Staley Road, Grand Island, New York, USA, 14072

Full list of author information is available at the end of the article

Results and discussion

The use of components A and B with CHO-S[®] cells in CD FortiCHO[™] medium causes a considerable increase in the level of galactosylation of the recombinant IgG (Figure 1) as shown by the shift in the glycosylation profile from G0F to G1F and G2F. The use of targeted transcriptome analysis revealed that the changes observed in the glycosylation profile do not translate to noticeable differences in the expression levels of the glycosylation genes. There are changes in gene expression levels with

culture age but they are not altered by the additions of components A and/or B. It was originally theorized that components A and B could act as cofactors or substrates within the glycosylation enzymatic pathways but this could not be confirmed without an understanding of the glycosylation gene profile. The changes in the glycosylation patterns combined with the absence of changes in the gene expression data lend support to this theory. With this information it is apparent that the additions of the glycosylation-enhancing components A and B can



increase galactosylation of recombinant proteins with no negative effect on growth, titer or glycosylation gene expression.

The comparison between CHO-S[®] and DG44 cultures without supplementation with components A or B revealed the DG44 culture had better galactosylation with increased proportions of G1F and G2F. Both cell lines express high levels of DDOST, RPN1, DAD1 and SST3A which are all part of the oligosaccharyltransferase complex which catalyzes the transfer of high mannose oligosaccharides from lipid-linked oligosaccharide donors to the asparagines on the Asn-X-Ser/Thr of the polypeptide chain. The DG44 cells differ from the CHO-S[®] cells with increases in: ALG2, ALG3, ALG9 and ALG12 (mannosyltransferases), ALG8 and ALG10 (glucosyltransferases), ALG14 (acetylglucosaminyltransferase), and B4GALT5 (galactosyltransferase). These increases in gene expression in DG44 cells seem to coincide with the higher galactosylation profiles observed in the glycan analysis.

Conclusions

Differences in growth, titer and glycoform distribution were observed between CHO-S[®] and CHO DG44 cells. DG44 cells had higher expression of glycosylation transferase genes compared to CHO-S[®] cells. Components A and B had synergistic effects on terminal galactosylation (Figure 1), showed no changes in gene expression and could be acting as cofactors/substrates with glycosylation enzymes.

Acknowledgements

The Austin team (Natalie Hernandez, Laura Chapman, Angie Cheng, Lea Kristi and Daniel Williams) for library preparation and transcriptome analysis.

Authors' details

¹Life Technologies Corporation, 3175 Staley Road, Grand Island, New York, USA, 14072. ²Life Technologies Corporation, 2130 Woodward, Austin, Texas, USA, 78744. ³Life Technologies Corporation, 29851 Willow Creek, Eugene, Oregon, USA, 97402.

Published: 4 December 2013

doi:10.1186/1753-6561-7-S6-P99

Cite this article as: Boniface et al.: Profiling of glycosylation gene expression in CHO fed-batch cultures in response to glycosylation-enhancing medium components. *BMC Proceedings* 2013 **7**(Suppl 6):P99.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

